

Measurement of the temperature-dependent speed of sound and change in Grüneisen parameter of tissue-mimicking materials

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Abstract—Knowledge of the temperature dependence of the material properties of tissue-mimicking materials is useful or essential for many applications. This includes photoacoustic thermometry where the temperature dependence of the Grüneisen parameter of tissues leads to changes in the recorded photoacoustic signal amplitude with temperature. Here, a setup is described that can measure the temperature dependence of the speed of sound and photoacoustic conversion efficiency ($\mu_a\Gamma$) of tissue-mimicking materials. Agar-based phantoms, copolymer-in-oil, gel wax, PVCP, silicone and water were characterised in the newly developed setup for temperatures between 22 °C and 50 °C. This information provides a valuable resource for material characterisation and future development of tissue-mimicking materials.

Index Terms—metrology, phantoms, photoacoustics, thermometry, ultrasound

I. Introduction

Photoacoustic (PA) thermometry is a rapidly emerging technique for non-invasive temperature monitoring that exploits the temperature dependence of the Grüneisen parameter Γ of tissues. This dependence leads to changes in the recorded photoacoustic signal amplitude with temperature and can thus be used to determine the temperature of that tissue [1]. In order to assess the accuracy, robustness and applicability of photoacoustic thermometry in different experimental scenarios, a stable phantom material with known properties is needed. This can also be used to tackle other challenges encountered in new imaging and therapeutic modalities, such as quality assurance [2]. However, current literature lacks information on the temperature dependency of the relevant phantom properties, such as the speed of sound and the Grüneisen parameter [3].

In this paper, we investigate the temperature-dependent speed of sound and Grüneisen parameter for some commonly used tissue-mimicking materials (TMMs) for photoacoustic and ultrasound imaging, namely agar-based phantoms, copolymer-in-oil, gel wax, PVCP, silicone, and water. For this purpose, a newly developed measurement

setup is presented, capable of measuring the desired properties between 22 and 50 °C.

II. Materials and Methods

A. Tissue-mimicking materials

The tissue-mimicking materials (TMMs) included in the study were agar-based materials [4], [5], copolymer-in-oil [6], gel wax [7], polyvinyl chloride plastisol (PVCP) [8] and silicone [9]. For each TMM, planar samples of 10 ± 5 mm thickness and 60 mm diameter were prepared with an embedded T-type thermocouple (5TC-TT-TI-36-1M-SMP-M IEC PFA-insulated, Omega Engineering Limited, Manchester, U.K.). The fabrication protocols were adopted from the literature, however, for the purpose of this study, acoustic scatterers were omitted and specialised black dyes were added to the base material in order to improve the signal-to-noise ratio. A summary of the base materials and dyes used in the fabrication of the phantoms, along with their relevant acoustic properties can be found in Table I.

B. Experimental setup

The experimental setup (Fig. 1) was comprised of a 1064 nm Q-Switched Nd:YAG laser (Ultra, Big Sky Laser Technologies, Bozeman, MT, USA) for illumination, a temperature-controlled water bath and a broadband PVDF receiver with a 5 mm active element and centre frequency of 14 MHz (PA1075, Precision Acoustics Ltd., Dorchester, U.K.).

The laser light was delivered using an optical fibre with a numerical aperture of approximately 12° (1.a) and uniform illumination of the sample achieved using a lens (1.b), spreading the beam diameter to approximately 30 mm. The pulse duration, energy, and repetition frequency were 5.6 ns, 45 mJ, and 20 Hz, respectively. For safety purposes, the entire setup was placed in a light-absorbing enclosure. The water bath consisted of a stainless steel tank (2.c) with inner dimensions of 150 mm \times 120 mm \times 50 mm (height \times width \times length) and water heaters (2.d). The tank contained an optical window (31 mm \times 31 mm BK7 window 1/4 wave 50 mm SQ NIR II, Edmund Optics Ltd., York, U.K.) at the front and a

This work was supported in part by the Engineering and Physical Sciences Research Council, grant numbers EP/L020262/1 and EP/P008860/1.

TABLE I: Base materials and dyes used for the fabrication of the tissue-mimicking materials and their speed of sound c and density ρ as reported in the literature.

TMM	Ref.	Base	Dye	c (ms ⁻¹)	ρ (kgm ⁻³)
Agar	[4], [5]	78.83% water, 11.21% glycerol, 3% agar, 0.95% 3- μ m Al ₂ O ₃ , 0.88% 0.3- μ m Al ₂ O ₃	India ink (Pelikan Vertriebsgesellschaft mbH & Co. KG, Hannover, Germany)	1536.7 \pm 8.7	1050 \pm 10.0
Copolymer-in-oil	[6]	12% copolymer oil + 3% LDPE	Caligo safe wash relief ink (Cranfield Colours, Cwmbran, UK)	1459 @ 3 MHz	900 \pm 5.0
Gel wax	[7]	FF1 003 (Mindsets Online, Waltham Cross, UK)	Caligo safe wash relief ink (Cranfield Colours, Cwmbran, UK)	1445 \pm 2.7	850 \pm 5.0
PVCP	[8]	Lure Flex Firm (Lure Factors, Doncaster, UK)	Black Plastic Color (Lure Factors, Doncaster, UK)	1400 @ 3 MHz	1000 \pm 5.0
Silicone	[9]	Polytek PlatSil SiliGlass (MB Fibreglass, Newtownabbey, UK)	Polycraft Black Silicone Pigment (MB Fibreglass, Newtownabbey, UK)	1030 @ 7 MHz	1070 \pm 30

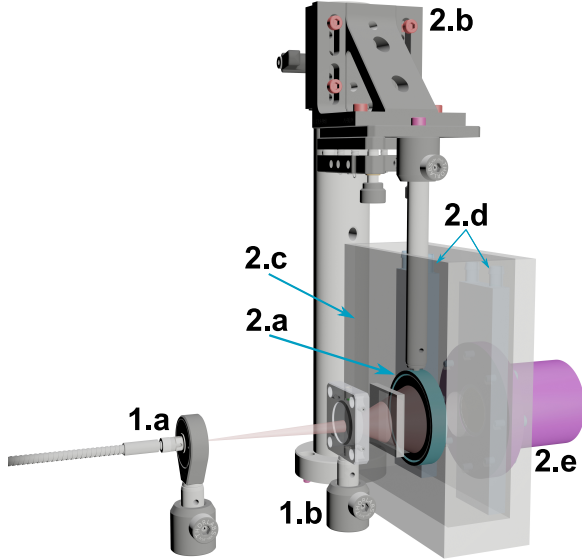


Fig. 1: Experimental setup comprising of a 1064 nm Ultra laser for illumination (1) and a temperature-controlled water bath (2). The light was delivered using an optical fibre (1.a) and uniform illumination of the sample achieved using a lens (1.b). The sample was held in a 3D printed holder (2.a), mounted to an optical post assembly with a 2-axis tilt (2.b) used for alignment and immersed in the water tank (2.c) with heaters (2.d). The generated photoacoustic signals were acquired using a broadband PVDF receiver with a 5 mm active element, which was held in a 3D printed transducer holder (2.e).

circular opening ($\varnothing = 23$ mm) for the transducer at the back. For the measurements, the tank was filled with degassed deionised water. The sample was placed in a circular 3D printed sample holder (2.a) and mounted to an optical post assembly with a 2-axis tilt (2.b) used for alignment. The sample was immersed in the water and aligned for maximum signal.

The generated photoacoustic plane waves were recorded using the PVDF receiver which was held in an SLS nylon 3D printed holder (2.e). The holder attached to the tank from the rear side using eight hex socket cap screws and a silicone gasket providing a leak-proof attachment. The PVDF receiver was connected to an oscilloscope (DSO-X 3024A, Agilent Technologies, PaloAlto, CA, USA) using a submersible hydrophone preamplifier that buffers the signal and provides a 50 Ohm source, which is powered by a DC coupler (Precision Acoustics Ltd., Dorchester, U.K.). Signals were digitised using the oscilloscope with a sampling frequency of 200 MHz and 128 averages, and acquired every 1 degree between 22 and 50 degrees Celsius during both heating and cooling. The experiments were repeated 5 times in order to characterise the repeatability of the measurements.

The water (and thus sample) heating was achieved using a thermostat with an external circulation (ECO RE415S Silver thermostat, Lauda Dr. R. Wobser GmbH, Lauda-Königshofen, Germany) connected to two aluminium heat sinks immersed in the tank (2.d). An application for control of the various parts of the measurement process was built using LabVIEW (National Instruments, Austin, TX, USA). The temperature control was implemented using LabVIEW's in-built PID controller virtual instrument. The communication between the application and the external instruments was made via a USB interface.

In order to verify the acquired data for the TMMs and characterise the newly developed experimental setup, measurements were repeated using degassed deionised water as a reference material. For this purpose, the sample holder was removed from the bath and an OPO system (SpitLight 600, Innolas, Krailling, Germany) providing 6

ns pulses at 30 Hz tuned to the water absorption peak at 1470 nm was used for the generation of photoacoustic signals.

C. Data analysis

The data acquired using the setup described above was analysed offline using a Matlab script (R2018a, MathWorks, Massachusetts, USA). The recorded signals for each sample and temperature were first averaged and their DC offset removed. The peak values of the averaged signals were then extracted, and a time-of-flight method used to calculate the speed of sound in the samples.

a) Bulk speed of sound: The temperature-dependent bulk speed of sound (group velocity) of the samples was determined using the time-of-flight method [10]. The time between the laser trigger and the maximum peak in the photoacoustic signal was used in conjunction with the known temperature-dependent speed of sound in distilled water [11]. The speed of sound of the sample (c_{sample}) was then calculated using the relation:

$$c_{\text{sample}} = d_{\text{sample}} \left(t - \frac{d_{\text{water}}}{c_{\text{water}}} \right)^{-1} \quad (1)$$

where d_{sample} is the sample thickness, t is the time of arrival of the photoacoustic signal, c_{water} is the propagation speed in water and d_{water} the water path length, ie. the distance between the sample and the receiver. The thickness of the samples was measured using a digital caliper with a resolution of ± 0.01 mm (Mitutoyo, Tokyo, Japan). The measurements were repeated five times per sample with a maximum standard deviation of 0.1 mm. For water measurements, Eq. (1) reduces to $c_{\text{water}} = \frac{d_{\text{water}}}{t}$ where d_{water} was taken to be equal to the length of the water tank.

b) Photoacoustic conversion efficiency: The photoacoustic conversion efficiency is the product of the optical absorption coefficient μ_a and the Grüneisen parameter Γ [12]. This efficiency is reported instead of the Grüneisen parameter as the temperature dependence of the optical absorption coefficient for most materials is not known. Thus the recorded photoacoustic signals were used to retrieve the temperature-dependent change in $\mu_a \Gamma$ by extracting the peak values of the averaged signals and plotting against the thermocouple temperature data.

III. Results and Discussion

The temperature-dependent speed of sound of water, agar-based phantoms, copolymer-in-oil, gel wax, PVCP and silicone phantoms are presented in Fig. 2.

It can be seen that the measured temperature-dependent speed of sound of water shows a consistent offset of approximately 5 ms^{-1} , while for the solid samples the values measured at the baseline temperature of the measurement are similar to the literature data for the used TMMs (Table I). These discrepancies can be contributed mostly to the uncertainty in sample thickness

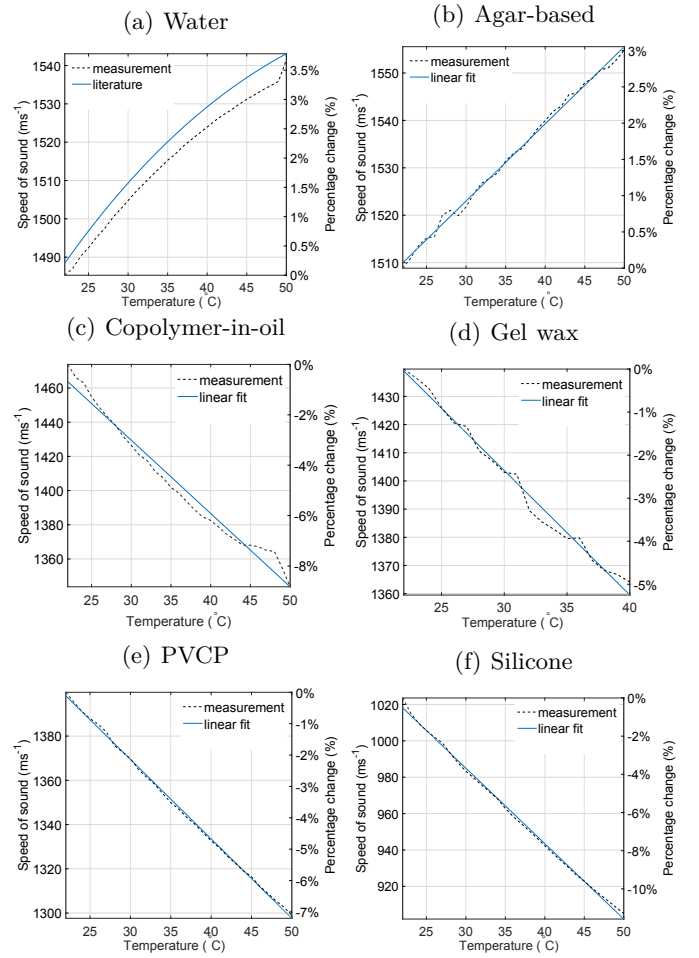


Fig. 2: Temperature-dependent speed of sound of (a) water, (b) agar-based phantoms, (c) copolymer-in-oil, (d) gel wax, (e) PVCP and (f) silicone phantoms.

and water path length measured using a caliper. In order to reduce these errors, future studies should employ a broadband through-transmission method [13] to evaluate sample thickness by using the reflected signals from the front and rear surface of the samples.

The changes in the PA conversion efficiency of the tissue-mimicking samples and water over the temperature range from 22°C to 50°C are presented in Fig. 3. The exception is gel wax, for which the maximum measurement temperature was 40°C due to its low melting point compared to the other materials. The results are an average of five measurement repeats and are normalised to the amplitude at 22°C . The error bars are omitted for clarity, with coefficients of variation $<1.5\%$ for all tested materials. The literature data for the temperature-dependent Grüneisen parameter of water is presented for reference, and was calculated using the approximation $\Gamma(T) = 0.0053T + 0.0043$ derived from the definition of

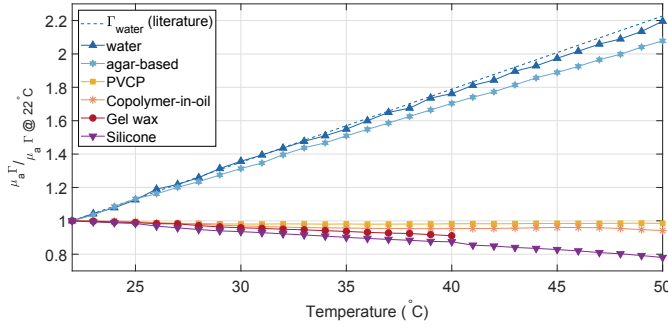


Fig. 3: The normalised temperature-dependent change in photoacoustic conversion efficiency $\mu_a \Gamma$ generated in phantoms over the temperature range from 22 to 50 °C. The literature data for the temperature-dependent Grüneisen parameter of water is presented for reference.

the Grüneisen parameter:

$$\Gamma = \frac{\beta c^2}{C_p} \quad (2)$$

and knowledge of the temperature dependence of the speed of sound, volume expansion thermal coefficient and specific heat under constant volume for water and aqueous solutions [14].

It can be seen that the measured temperature-dependent change in the photoacoustic conversion efficiency of water corresponds closely to the literature data for the Grüneisen parameter of water within the measurement temperature range, which increases by approximately 4% per degree Celsius.

The results for the TMMs can be sorted into three distinct categories. The first one is the agar-based phantoms. As their content is mostly water, the photoacoustic signal amplitudes generated in these samples increase with temperature accordingly. Copolymer-in-oil phantoms and PVCP, on the other hand, do not seem to exhibit any significant change in the PA conversion efficiency with temperature. Although their speed of sound was measured to decrease by 7-8% within the measurement range, the remaining material properties contributing to the Grüneisen parameter (Eq. (2)) and their dependency on temperature are unknown. The change in the PA conversion efficiency in gel wax and silicone phantoms exhibits a decreasing trend with temperature. This is expected due to their oil content arising from the base material and dye used, for which the speed of sound is known to decrease with temperature [15].

IV. Conclusion

The temperature-dependent speed of sound and change in photoacoustic conversion efficiency for selected tissue-mimicking materials is presented. This information forms a valuable resource for the future development of TMMs with properties suitable for applications in photoacoustic thermometry and photoacoustic imaging in general. These

measurements also have the potential to provide indirect information about the thermodynamics of the materials.

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